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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:) Interference No: 102,822
)
MARC ALIZON et al.)
)
Serial No.: 158,652) Group Art Unit: 1805
)
Filed: February 22, 1988) Examiner: J. Railey

For: CLONED DNA SEQUENCES RELATED TO THE GENOMIC RNA OF LYMPHADENOPATHY ASSOCIATED VIRUS (LAV) AND PROTEINS ENCODED BY SAID LAV GENOMIC RNA

Honorable Commissioner of Patents
and Trademarks
Washington, D.C. 20231

Sir:

CLAIM FOR PRIORITY

Under the provisions of 35 U.S.C. §119, applicants hereby claim the benefit of the filing date of British Application No. 84 23659, filed September 19, 1984, for the above-identified United States patent application.

In support of applicants' claim for priority, filed herewith is one certified copy of the above.

Respectfully submitted,

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Dated: June 21, 1993

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CERTIFICATE OF SERVICE

It is hereby certified that a true copy of CLAIM FOR PRIORITY WITH CERTIFIED COPY OF UNITED KINGDOM APPLICATION NO. 84 23659 ATTACHED is being sent via first class mail, postage prepaid to Guy W. Chambers, Townsend and Townsend, Steuart Street Tower, 20th Floor, One Market Plaza, San Francisco, CA 94105 this 21st day of June 1993.

Dated: June 21, 1993 By: Jean B. Fordi

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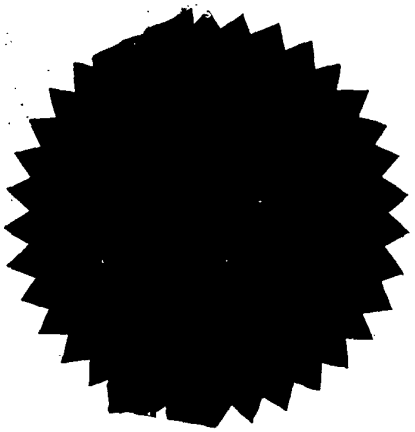

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PATENTS ACT 1977

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PATENTS FORM No. 1/77 (Revised 1982)
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REQUEST FOR GRANT OF A PATENT

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THE GRANT OF A PATENT IS REQUESTED BY THE UNDERSIGNED ON THE BASIS OF THE PRESENT APPLICATION

I Agent's Reference JJD/EAF/26679

II Title of Invention CLONED DNA SEQUENCES, HYBRIDIZABLE WITH GENOMIC RNA OF LYMPHADENOPATHY-ASSOCIATED VIRUS (LAV)

III Applicant or Applicants *(See note 2)*

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(a) The applicant(s) is/are the sole/joint inventor(s)
or
(b) A statement on Patents Form No. 7/77 is/will be furnished

V Name of Agent (if any) *(See note 4)* Reddie & Grose | ADP CODE NO

VI Address for Service (See note 5) 16 Theobalds Road
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VII Declaration of Priority (See note 6)

Country	Filing date	File number

VIII The Application claims an earlier date under Section 8(3), 12(6), 15(4), or 37(4) (See note 7)

Section No.

Earlier application or patent number and filing date

IX Check List *(To be filled in by applicant or agent)*

A The application contains the following number of sheet(s)			B The application as filed is accompanied by:–	
1	Request	1 Sheet(s)	1	Priority document NO
2	Description	18 Sheet(s)	2	Translation of priority document NO
3	Claim(s)	3 Sheet(s)	3	Request for Search NO
4	Drawing(s)	2 Sheet(s)	4	Statement of Inventorship and Right to Apply NO
5	Abstract	0 Sheet(s)	5	

X It is suggested that Figure No 1 of the drawings (if any) should accompany the abstract when published.

XI Signature *(See note 8)*

Reddie & Grose, Agents for the Applicant(s)

NOTES:

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2. Enter the name and address of each applicant. Names of individuals should be indicated in full and the surname or family name should be underlined. The names of all partners in a firm must be given in full. Bodies corporate should be designated by their corporate name and the country of incorporation and, where appropriate, the state of incorporation within that country should be entered where provided. Full corporate details, eg "a corporation organised and existing under the laws of the State of Delaware, United States of America," trading styles, eg "trading as xyz company", nationality, and former names, eg "formerly [known as] ABC Ltd." are not required and should not be given. Also enter applicant(s) ADP Code No. (if known).
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6. The declaration of priority at VII should state the date of the previous filing and the country in which it was made and indicate the file number, if available.
7. When an application is made by virtue of section 8(3), 12(6), 15(4), or 37(4) the appropriate section should be identified at VIII and the number of the earlier application or any patent granted thereon identified.
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Cloned DNA sequences, hybridizable with genomic RNA of lymphadenopathy-associated virus (LAV)

The invention relates to cloned DNA sequences hybridizable TOgenomic RNA and DNA of lymphadenopathy-associated virus (LAV), a process for their preparation and their uses. It relates more particularly to stable probes including a DNA sequence which can be used for the detection of the LAV virus or related viruses or DNA proviruses in any medium, particularly biological, samples containing of any them.

Lymphadenopathy-associated virus (LAV) is a human retrovirus first isolated from the lymph node of a homosexual patient with lymphadenopathy syndrome, frequently a prodrome or a benign form of acquired immune deficiency syndrome (AIDS) (cf.1). Subsequently other LAV isolates have been recovered from patients with AIDS or pre-AIDS (cf. 2-5). All available data are consistent with the virus being the causative agent of AIDS (cf. 11).

The virus is propagated on activated T lymphocytes and has a tropism for the T-cell subset OKT4 (cf. 2-6), in which it induces a cytopathic effect. However, it has been adapted for growth in some Epstein-Barr virus transformed B-cell lines (cf. 7), as well as in the established T-lymphoblastic cell line, CEM.

LAV-like viruses have more recently been independently isolated from patients with AIDS and pre-AIDS. These viruses called HTLV-III (Human T-cell Leukemia/Lymphoma virus type III (cf. 12-15) and ARV (AIDS-associated retrovirus) seem to have many characteristics similar to those of LAV and it is thus probable that they represent independent isolates of the LAV prototype.

Detection methods so far available are based on the recognition of core proteins. Such a method is

disclosed in European application titled "Antigènes, moyens et méthode pour le diagnostic de lymphadénopathie et du syndrome d'immunodépression acquise" filed on September 14, 1984 under the priority of British application Serial Nr. 83 24000 filed on September 15, 1983. As a matter of fact a high prevalence of anti-p25 antibodies has been found in the sera of AIDS and pre-AIDS patients and to a lower but significant extent in the high-risk groups for AIDS (cf. 8-10). However the same sera were found not to recognize the virus as a whole, in a non-disintegrated state.

The present invention aims at providing new means which should not only also be useful for the detection of LAV or related viruses (hereafter more generally referred to as "LAV viruses"), but also have more versatility, particularly in detecting specific parts of the genomic DNA of said viruses whose expression products are not always detectable by immunological methods.

The DNAs according to the invention consist of DNAs which contain DNA fragments, hybridizable with the genomic RNA of LAV. Particularly said DNAs consist of said cDNAs or cDNA fragments or of recombinant DNAs containing said cDNAs or cDNA fragments.

Preferred clonedcDNA fragments respectively contain the following restriction sites in the respective orders which follow (from the 3' end to the 5' end) :

1) HindIII, SacI, BglII (LAV75)
2) HindIII, SacI, BglII, BglII, KpnI (LAV82)
3) HindIII, SacI, BglII, BglII, KpnI, XhoI, BamHI, HindIII, BglII (LAV13).

The LAV75, LAV82 and LAV13 designations correspond to the designations of the recombinant plasmids designated as pLAV 75, pLAV 82 and pLAV 13 respectively, in which they were first cloned. In other words LAV 75, LAV 82 and LAV 13 respectively present as inserts in said recombinant plasmids. For convenience the designations LAV 75, LAV 82

and LAV 13 will be further used throughout this specification to designate the cDNA fragments, whether the latter are in isolated form or in a plasmid forms, whereby the other DNA parts of said last mentioned recombinants are identical to or different of the corresponding parts of pLAV 75, pLAV 82 and pLAV 13 respectively.

Preferred cDNAs also (like LAV 75, LAV 82 and LAV 13) contain a region corresponding to the R and U 3 regions of the LTR (Long Terminal Repeat) as well as the 3' end of the coding region of the retroviral DNA. Particularly if it is assumed that the retroviral structure of LAV is in general agreement with the retroviral genomic structures to date.

LAV 13 which has a size of about 2.5 Kbp has been found of particular advantage. It is highly specific of LAV or LAV related viruses and does also recognizes more of the LAV retroviral genomes than do LAV75 or LAV82. Particularly LAV 13 enabled the identification of the RU 5 junction of the retroviral genomes within the LTR and, subsequently, the sizes of the LAV genomes, which average from about 9.1 to about 9.2 kb.

LAV 13 is free of restriction sites for the following enzymes Eco RI, Nru I, Pvu I, Sal I, Sma I, Sph I, Stu I and Xba I.

LAV 13 further appears to contain at least part of the DNA sequences corresponding to those which, in retroviral genomes, code for the envelope protein.

The invention further relates to any of the fragments contained in the cDNA which seems to correspond to part of the whole of the LAV retroviral genome, which is characterized by a series of restriction sites in the order hereafter (from the 5' end to the 3' end).

The coordinates of the successive sites of the whole LAV genome (restriction map) are indicated hereafter too, with respect to the Hind III site (selected as of coordinate 1) which is located in the R region. The

coordinates are estimated to within ± 200 bp. Some coordinates are better established than others.

Hind III	0
Sac I	50
Bam HI	460
Hind III	520
Bam HI	600
Pst I	800
Hind III	1 100
Bgl II	1 500
Kpn I	3 500
Kpn I	3 900
Eco RI	4 100
Eco RI	5 300
Sal I	5 500
Kpn I	6 100
Bgl II	6 500
Bgl II	7 600
Hind III	7 850
Bam HI	8 150
Xho I	8 600
Kpn I	8 700
Bgl II	8 750
Bgl II	9 150
Sac I	9 200
Hind III	9 250

The abovesaid DNA according to the invention optionally contains an additional Hind III approximately at the 5 550 coordinate.

The invention further relates to other preferred DNA fragments corresponding substantially to those which in relation to the abovesaid restriction map extend respectively :

- from approximately Kpn I (6 100) to approximately Bgl II (9150) said fragment being thought to correspond at least in part to the gene coding for the proteins of the

envelope ; in particular a protein p110 of about 110,000 Daltons is encoded by this region ;

- from approximately Kpn I (3 500) to approximately Bgl II (6500), said fragment being thought to correspond at least in part to the pol gene, coding for the virus polymerase ;
- from approximately Pst (800) to approximately Kpn I (3500), said fragment being thought to correspond at least in part to the gag gene, which codes for the core antigens, including the p25, the p18, and the p13 proteins.

More particularly the invention relates to any fragment corresponding to the above ones, having substantially the same sites at substantially same distances from one another, all of these fragments having in common the capability of hybridizing with the LAV retroviral genomes. It is of course understood that fragments which would include some deletions or mutation which would not substantially alter their capability of also hybridizing with the LAV retroviral genomes are to be considred as forming obvious equivalents of the DNA fragments more specifically referred to hereabove.

Additional features of the invention will appear in the course of the disclosure of additional features of preferred DNAs of the invention, the preparation conditions and the properties of which will be illustrated hereafter in a non limitative manner. Reference will also be had to the drawings in which :

- fig. 1 shows restriction maps of preferred LAV inserts contained in plasmid recombinants ;
- fig. 2 shows restriction maps of complete LAV fragments.

1. Construction of a cDNA library

1.1 Virus purification

Virions were purified from FR8, an immortalized, permanent LAV producing B-Lymphocyte line (cf. 7) (deposited at the "Collection Nationale de Cultures de Micro-organismes" of the INSTITUT PASTEUR of Paris, under Nr. I-303 on May 9, 1984). The purification protocol was

described(cf. 1). The main steps were : polyethylene-glycol treatment of culture supernatant, pelleting through 20 % sucrose cushion, banding on 20-60 % sucrose gradient and pelleting of the virus-containing fractions.

1.2 First-strand cDNA synthesis

The virus associated detergent activated endogenous reaction is a technique bringing into play the reverse transcriptase of the virus, after purification thereof and lysis of its envelope.

For each reaction, purified virus corresponding to 250-300 ml of FR8 supernatant was used. Final reaction volume was 1 ml. Incubation was at 37°C for 45 mn. Protein concentration was about 250 microg/ml. Buffer was : NaCl 25 mM ; Tris HCl pH 7.8 50 mM, dithiothreitol 10 mM, $MgCl_2$ 6 mM, each of dATP, dGTP, dTTP at 0.1 mM, Triton X-100 0.02 % ; oligo dT primer 50 microg/ml. The cDNA was labelled 15 mn with alpha ^{32}P-dCTP 400 Ci/mmole to 0.6 microM plus cold dCTP to 4 microM. Afterwards, cold dCTP was added to 25 microM to ensure optimal elongation of the first strand.

The reaction was stopped 30 mn after the dCTP chase by adding EDTA to 20 mM, SDS to 0.5 %, digesting one hour with proteinase K at 100 microg/ml and phenol-chloroform extraction.

cDNA was then purified on G-50 Sephadex (Pharmacia) and ethanol precipitated.

1.3 2nd strand synthesis and cloning

Purified cDNA-RNA hybrids were treated with DNA polymerase I and RNase H, according to GUBLER and HOFFMAN (cf. 17). Double-stranded cDNA was dC-tailed with terminal transferase and annealed to dG-tailed Pst-digested pBR 327 (cf. 34) a derivative of pBR 322.

A cDNA library was obtained by transfection of *E. coli* C 600 recBC strain.

2. Detection of LAV-specific clones

2.1 Screening of the library

500 recombinant clones were grown on nitrocellulose filtres and *in situ* colony hybridization (cf. 35) was performed with another batch of cDNA made in endogenous virus-associated reaction as described (cf. 1.2) and labelled with ^{32}P. About 10 % of the clones could be detected.

A major family was obtained by small-scale amplification of these clones and cross-hybridization of their inserts. Among these clones a major family of hybridizing recombinants was identified. Three of these cDNA clones, named pLAV 13, 75 and 82, carrying inserts of 2.5, 0.6 and 0.8 kb respectively were further characterized (fig. 1).

All three inserts have a common restriction pattern at one end, indicating a common priming site. The 50 bp long common Hind III-Pst I fragment was sequenced (fig. 1) and shown to contain a polyA stretch preceeding the cloning dC tail. The clones are thus copies of the 3' end of a polyA-RNA.

The LAV 13 specificity was shown by different assays.

The specificity of pLAV 13 was determined in a series of filter hybridization experiments using nick-translated pLAV 13 as a probe. Firstly the probe hybridized to purified LAV genomic RNA by dot and Northern blotting (data not shown). pLAV 13 also hybridizes to the genomic RNA of virus concentrated from culture supernatant directly immobilized on filters (dot blot technique). LAV RNA from different sources : normal T-cells, FRG and other B-cell LAV producing lines, CEM cells and, although less strongly, LAV from the bone marrow culture from a haemophiliac with AIDS (cf. 3) were detected in a similar manner. Uninfected cultures proved negative. This rapid dot blot technique can be adapted with minor modifications

to the detection of LAV in serum or other body fluids.

Secondly the probe detected DNA in the Southern blots of LAV-infected T-lymphocytes and in the LAV-producing CEM cell line. No hybridization was detected in the DNA of uninfected lymphocytes nor in the DNA from normal liver (data not shown) under the same hybridization conditions.

A third characteristic resulted from the possibility of using LAV 13 to identify the whole retroviral genome of the LAV viruses as disclosed hereafter. Particularly characteristic 1.45 kb Hind III fragment which co-migrates with an internal viral fragment in Hind III cleaved pLAV 13 was detected. Bands at 2.3 and 6.7 kb were also detected. As the probe was only 2.5 kb long and as no junction fragments could be detected, it is probable that these extra-bands represent internal fragments arising from a Hind III polymorphism of the LAV genome.

Together these data show that pLAV 13 DNA is exogenous to the human genome and detects both RNA and integrated DNA forms derived from LAV infected cells. Thus pLAV 13 is LAV specific. Being oligo-dt primed, pLAV 13 must contain the R and U3 regions of the LTR as well as the 3' end of the coding region, assuming a conventional retroviral genome structure.

Cloning of LAV genomic DNA

Having found a HindIII site within the R region of the LTR, it was decided to clone the LAV genome by making a partial Hind III digest of proviral DNA from LAV infected cells. It was found that : (a) partial digestion increased the chance of isolating complete clones and (b) Hind III fragments were easily cloned in lambda replacement vectors. The DNA isolated from T-cells of a healthy donor after *in vitro* infection with LAV was partially digested with Hind III and fractionated. A 9^{+} 1.5 kb DNA containing fraction was precipitated and ligated into the Hind III arms of lambda-L47.1 (cf. 18).

The cloning of LAV genomic DNA was carried out more particularly as follows :

cDNAs was prepared from LAV infected T cells as described above, then partially digested with Hind III and fractionated on a 5-40 % sucrose gradient in 10 mM Tris.Cl pH 8, 10 mM EDTA, 1 M NaCl (SW41 rotor, 16 hours at 40 000 rpm). A single fraction (9 $\pm$ 0.5 kb) was precipitated with 20 microg/ml Dextran T40 as carrier and taken up in TE-buffer (10 mM Tris.Cl pH 8, 1 mM EDTA). Lambda-L47.1 Hind III arms were prepared by frist ligating the cos sites followed by Hind III digestion and fractionation through a 5-40 % sucrose gradient. Fractions containing only the lambda-Hind III arms were pooled, precipitated and taken up in TE-buffer. Ligation of arms to DNA was made at approximately 200 microg DNA/ml using a 3:1 molar excess of arms and 300 units of T4 DNA ligase (Biolabs). In vitro packaging lysates were made according to (38). After in vitro packaging the phage lysate was plated out on NM538 on a C600 recBC strain. Approximately two million plaques were screened by in situ hybridization (cf. 39) using nitrocellulose filters. Hybridization was performed at 68°C in 1 x Denhardt solution, 0.5 % SDS, 2 x SSC, 2 mM EDTA. Probe : ^{32}P nick-translated LAV insert of pLAV 13 at $>10^8$ cpm/microg : Filters were washed 2 x 30 minutes in 0-1 SSC, 0.1 % SDS at 60°C, and exposed to Kodak XAR-5 film for 29-40 hours. Seven positive clones were identified and plaque purified on a C 600 rec BC strain. Liquid cultures were grown and the recombinant phages banded in CsCl. Plage DNA was extracted and digested under the appropriate conditions.

Seven independent clones were so derived from approximatively two million phage plaques after screening in situ with a nick-translated pLAV 13 insert as a probe. Restriction maps of lambda-J19 as well as of a Hind III polymorph lambda-J61 are shown in fig. 2. Other recombinants lambda-J27, lambda-J31 and lambda-J57 had the same

Hind III map as lambda-J19. The map of lambda-J81 is identical but for an additional Hind III site at coordinate of approximately 5 550.

The restriction maps of fig. 2 were oriented by hybridizing blots with respect to pLAV 13 DNA.

The restriction map of the LAV 13 cDNA clone is also shown in fig. 2. The restriction sites of lambda-J19 are : B-Bam HI, Bg-Bgl II, H-Hind III, K-Kpn I, P-Pst I, R-Eco RI, S-Sac I, Sa-Sal-I and X-Xho I. Underneath the scale is a schema for the general structure of the retroviruses showing the LTR elements U3, R and U5. Only the R/US boundary has been defined and other boundaries are only drawn figuratively.

There may be other Bam HI sites in the 5' 0.52 kb Hind III fragment of lambda-J19. They generate fragments that are too small to be detected.

Fig. 2 also shows those Hind III fragments of lambda-J19 and lambda-J81 which are detected by pLAV 13 (marked (+)), those which are not detected (-).

More particularly lambda-J19 shows four Hind III bands of 6.7, 1.45, 0.6 and 0.52 kb the first two of which correspond to bands in the genomic blot of Hind III restricted DNA. The smallest bands of 0,6 and 0,52 kb were not seen in the genomic blot but the fact that they appear in all the independently derived clones analyzed indicates that they represent internal and not junction fragments, assuming a random integration of LAV proviral DNA. Indeed, the 0,5 kb band hybridizes with pLAV 13 DNA (fig. 2) through the small Hind III-Pst I fragment of pLAV 13. Thus the 0,5 kb Hind III fragment of lambda-J19 contains the R-U5 junction within the LTR.

It appears that lambda-J81 is a restriction site polymorph of lambda-J19. Lambda-J81 shows five Hind III bands of 4.3, 2.3, 1.45, 0.6 and 0.52 kb. The 2.3 kb band is readily detected in the genomic blot by a pLAV 13 probe, but not the 4.3 kb fragment. That lambda-J81 is a

Hind III polymorph and not a recombinant virus is shown by the fact that nick-translated lambda-J19 DNA hybridizes to all five Hind III bands of lambda-J81 under stringent hybridization and washing conditions. Also other restrictions sites in lambda-J81 are identical to those of lambda-J19.

Relationship to other human retroviruses

HTLV-I and HTLV-II constitute a pair of C-type transforming retroviruses with a tropism for the T-cell subset, OKT4 (cf. 20). An isolate of HTLV-I has been totally sequenced (cf. 21) and partial sequencing of an HTLV-II has been reported (cf. 22-24). Both genomes (one LTR) were approximately 8.3 kb in length, have a pX region and show extensive sequence homology. They hybridize between themselves under reasonably stringent conditions (40 % formamide, 5 XSSC) and even at 60 % formamide the pX regions hybridize (cf. 26). Thus a conserved pX region is a hallmark of this class of virus.

We have compared cloned LAV DNA and cloned HTLV-II DNA (pMO (cf. 27)) by blot-hybridization and found no cross-hybridization under low stringency conditions of hybridization and washing. For example, Hind III digested lambda-J19, lambda-J27 and lambda-J81 were electrophoresed, blotted and hybridized overnight with ^{32}P nick-translated pMO (HTLV-II) DNA (having a specific activity greater than 0.5 X 10^8 cpm/microg) in 20 % formamide, 5 XSSC, 1 X Denhardts solution, 10 % Dextran sulphate, at 37°C. The washings were repeated at 50°C and 65°C ~~65°C with~~ filters were washed at 37°C (t_m.50) t_m.50 using a 53.1 % GC content derived from the HTLV-I sequence (21) ~~referred~~ in 1 x SSX, 0.1 % SDS. Even when hybridized in 20 % formamide, 8 X SSC (t_m.50) and washed at 37°C in 2 X SSC (t_m.50) no hybridization was detected after two days exposure at -70°C using an intensifying screen.

Thus there is no molecular evidence of a relationship between LAV and the HTLV viruses. In addition, the

LAV genome is approximately 9 kb long in contrast to 8.3 kb for the HTLV viruses. Despite their comparable genome sizes LAV and Visna (cf. 29) cloned viral genomes do not cross-hybridize, nor does LAV with a number of human endogenous viral genomes (cf.30) under non stringent conditions (hybridization-20 % formamide, 8 SSC, 37°C ; washing - 2 SSC, 0.1 % SDS, 37°C.

The invention also relates more specifically to cloned probes which can be made starting from any DNA fragment according to the invention, thus to recombinant DNAs containing such fragments, particularly any plasmids amplifiable in procaryotic or eucaryotic cells and carrying said fragments. As mentioned earlier a preferred DNA fragment is LAV 13.

Using the cloned provirus DNA as a molecular hybridization probe - either by marking with radionucleotides or with fluorescent reagents - LAV virion RNA may be detected directly in the blood, body fluids and blood products (e.g. of the antihemophylic factors such as Factor VIII concentrates) and vaccines, i.e. hepatitis B vaccineIt has alrady been shown that whole virus can be detected in culture supernatants of LAV producing cells. A suitable method for achieving that detection comprises immobilizing virus onto said a support e.g. nitrocellulose filters, etc., disrupting the virion and hybridizing with labelled (radiolabelled or "cold" fluorescent- or enzyme-labelled) probes. Such an approach has already been developed for Hepatitis B virus in peripheral blood (according to SCOTTO J. et al. Hepatology (1983), 3, 379-384).

Probes according to the invention can also be used for rapid screening of genomic DNA derived from the tissue of patients with LAV related symptoms, to see if the proviral DNA or RNA is present in host tissue and other tissues.

A method which can be used for such screening

comprise the following steps : extraction of DNA from tissue,
restriction enzyme cleavage of said DNA, electrophoresis of the fragments and Southern blotting of genomic DNA from tissues, subsequent hybridization with labelled cloned LAV provival DNA. Hybridization in situ can also be used.

Lymphatic fluids and tissues and other non-lymphatic tissues of humans, primates and other mammalian species can also be screened to see if other evolutionnary related retrovirus exist. The methods referred to hereabove can be used, although hybridization and washings would be done under non stringent conditions.

The DNA according to the invention can be used also for achieving the expression of LAV viral antigens for diagnostic purposes as well as far the production of a vaccine against LAV. Of particular advantage in that respect are the DNA fragments coding core (gag region) and for envelope proteins, particularly the DNA fragment extending from Kpn I (6 100) to BglII(9 150).

The methods which can be used are multifold :

a) DNA can be transfected into mammalian cells with appropriate selection markers by a variety of techniques, calcium phosphate precipitation, polyethylene glycol, protoplast-fusion, etc..

b) DNA fragments corresponding to genes can be cloned into expression vectors for E. coli , yeast or mammalian cells and the resultant proteins purified.

c) The provival DNA can be "shot-gunned" (fragmented) into procaryotic expression vectors to generate fusion polypeptides. Recombinant producing antigenically competent fusion proteins can be identified by simply screening the recombinants with antibodies against LAV antigens .

d) The invention also relates to oligopeptides deduced from the DNA sequence of LAV antigen-genes to produce immunogens and antigens and which can be synthethised chemically.

All of the above (a-d) can be used in diagnostics as sources of immunogens or antigens free of viral particles, produced using non-permissive systems, and thus of little or no biohazard risk.

The invention further relates to the hosts (procaryotic or eucaryotic cells) which are transformed by the above mentioned recombinants and which are capable of expressing said DNA fragments.

Finally it also relates to vaccine compositions whose active principle is to be constituted by any of the expressed antigens, i.e. whole antigens, fusion polypeptides or oligopeptides.

The invention finally refers to the purified genomic mRNA, which can either be extracted as such from the LAV viruses or resynthesized back from the cDNA, particularly to a purified mRNA having a size approximating 9.1 to 9.2 kb, hybridizable to any of the DNA fragments defined hereabove or to parts of said purified mRNA. The invention also relates to parts of said RNA. The nucleotidic structures of this purified RNA or of the parts thereof can indeed be deduced from the nucleotidic sequences of the related cDNAs.

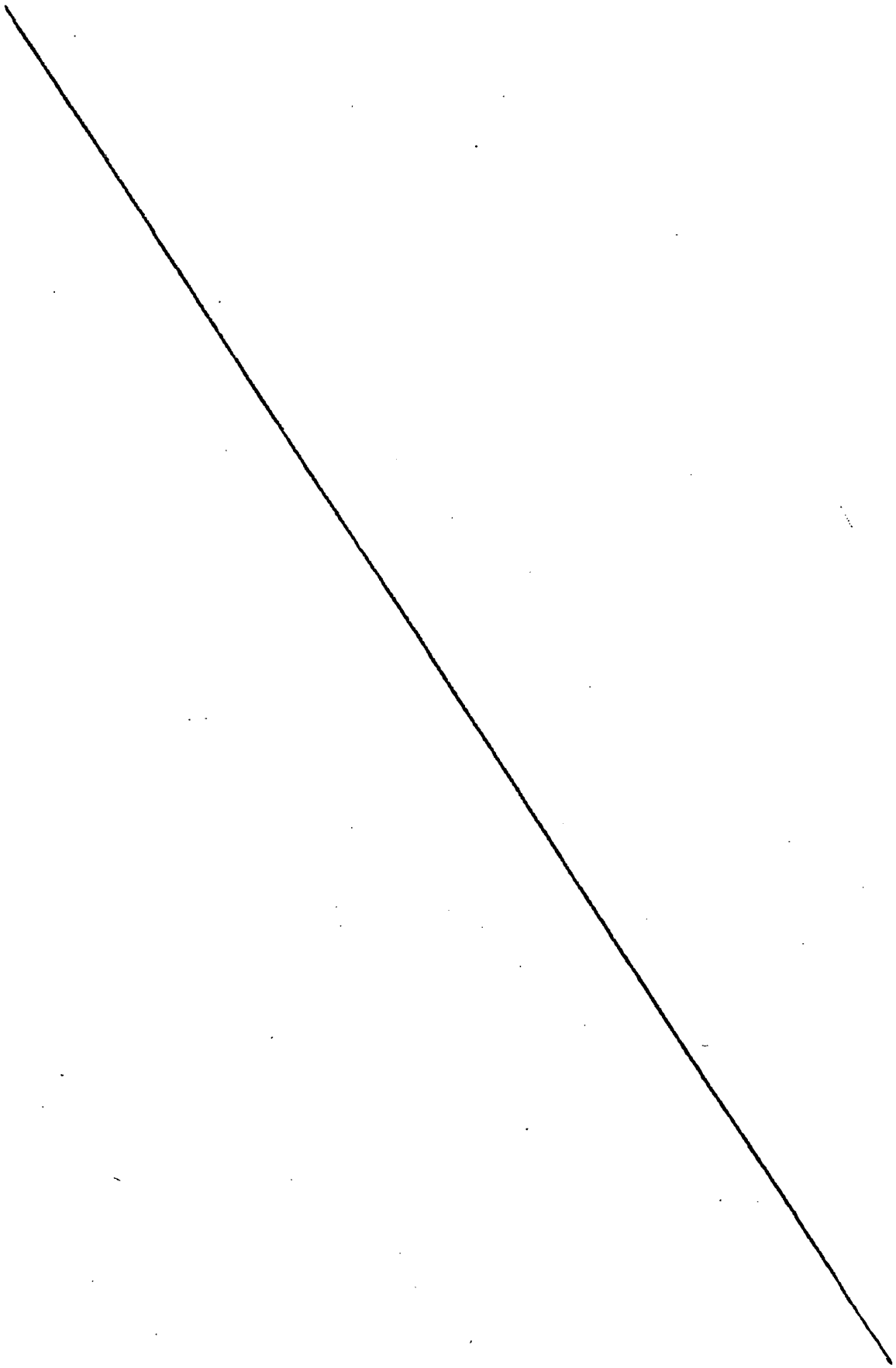
It will finally be mentioned that lambda-J19 and lambda-J81 have been deposited at the Collection Nationale des Cultures de Micro-organismes (C.N.C.M.) of the INSTITUT PASTEUR of Pasteur (France) under Nr. I-338 and I-339 respectively, on September 11, 1984.

The invention finally refers to the genomic DNA, the DNA sequence of which can be determined and used to predict the aminoacid sequences of the viral protein (antigens) and to the RNA probes which can be derived from the cDNA.

There follows the bibliography to which references have been made throughout this specification by bracketted numbers.

All the publications referred to in this

bibliography are incorporated herein by reference.



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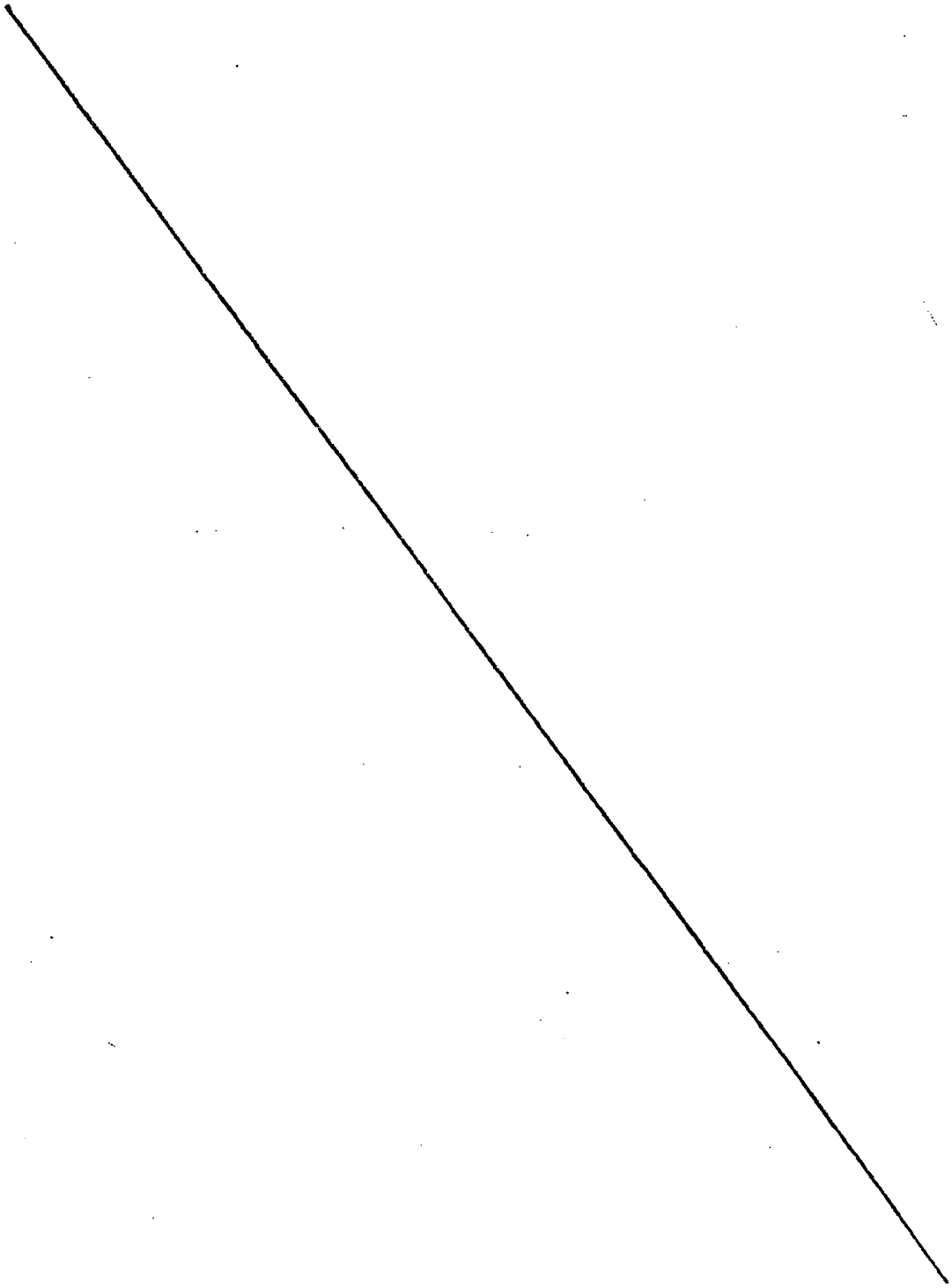

CLAIMS :

1. A cloned DNA which contains a DNA which is hybridizable with the genomic RNA of the LAV viruses or a fragment of said hybridizable DNA.

2. The DNA of claim 1 which is a recombinant of said hybridizable DNA or DNA fragment hybridizable with the genomic RNA of the LAV virus.

3. The DNA of claim 1 or 2 wherein said hybridizable DNA or DNA fragment is a cDNA.

4. The DNA of claims 1 to 3 which contains the following restriction sites in the following order (from the 3' end to the 5' end) :

Hind III, Sac I, Bgl II (LAV 75).

5. The DNA of claim 4 which contains the following restriction sites in the following order :

Hind III, Sac I, Bgl II, Bgl II, Kpn I (LAV 82).

6. The DNA of claim 4 which contains the following restriction sites in the following order :

Hind III, Sac I, Bgl II, Bgl II, Kpn I, XHo I, Bam HI, Hind III, Bgl II (LAV 13).

7. The DNA of claim 6 which has a size of about 2.5 kb.

8. The DNA of any of claims 1 to 7 which contains a region corresponding to the R and U3 regions of the LTR as well as to the 3' end of the coding region of the retroviral DNA.

9. The DNA of claim 1 which has a size from about 9.1 to 9.2 kb.

10. The DNA of claim 9 which contains the following series of restriction sites :

Hind III	0
Sac I	50
Bam HI	460
Hind III	520
Bam HI	600
Pst I	800

Hind III	1 100
Bgl II	1 500
Kpn I	3 500
Kpn I	3 900
Eco RI	4 100
Eco RI	5 300
Sal I	5 500
Kpn I	6 100
Bgl II	6 500
Bgl III	7 600
Hind III	7 850
Bam HI	8 150
Xho I	8 600
Kpn I	8 700
Bgl I	8 750
Bgl I	9 150
Sac I	9 200
Hind III	9 250

11. The DNA of claim 10 which contains an additional Hind III approximately at the 5 550 coordinate.

12. A DNA fragment according to claim 1 which comprises a sequence extending from approximately Kpn I (6100) to approximately Bam HI (8150) of the sequence defined in claim 11.

13. A DNA fragment according to claim 1 which comprises a sequence extending from approximately Kpn I (3500) to approximately Bgl II (6500) of the sequence defined in claim 11.

14. A DNA fragment according to claim 1 which comprises a sequence extending from approximately Pst (800) to approximately Kpn I (3500) of the sequence defined in claim 11.

15. A DNA fragment of claim 1 which codes for the enveloppe proteins.

16. A DNA fragment of claim 1 which codes for the retroviral polymerase.

17. A DNA fragment which codes for the core proteins.

18. A probe for the in vitro detection of LAV which consists of a DNA according to any of claims 1 to 17.

19. An expression vector, particularly a plasmid, for the transformation of procaryotic or eucaryotic cells which contains an insert consisting of a DNA fragment hybridizable with the retroviral genome of LAV viruses as defined in any of claims 1 to 17.

20. The vector of claim 18 which contains the DNA fragment of claim 15.

21. A microorganism, eucaryotic or procaryotic cell which is transformed by a vector according to claim 19 or 20 and which expresses the polypeptide encoded by the corresponding DNA fragment.

22. The purified RNAs of LAV viruses which have sizes from 9.1 to 9.2 kb.

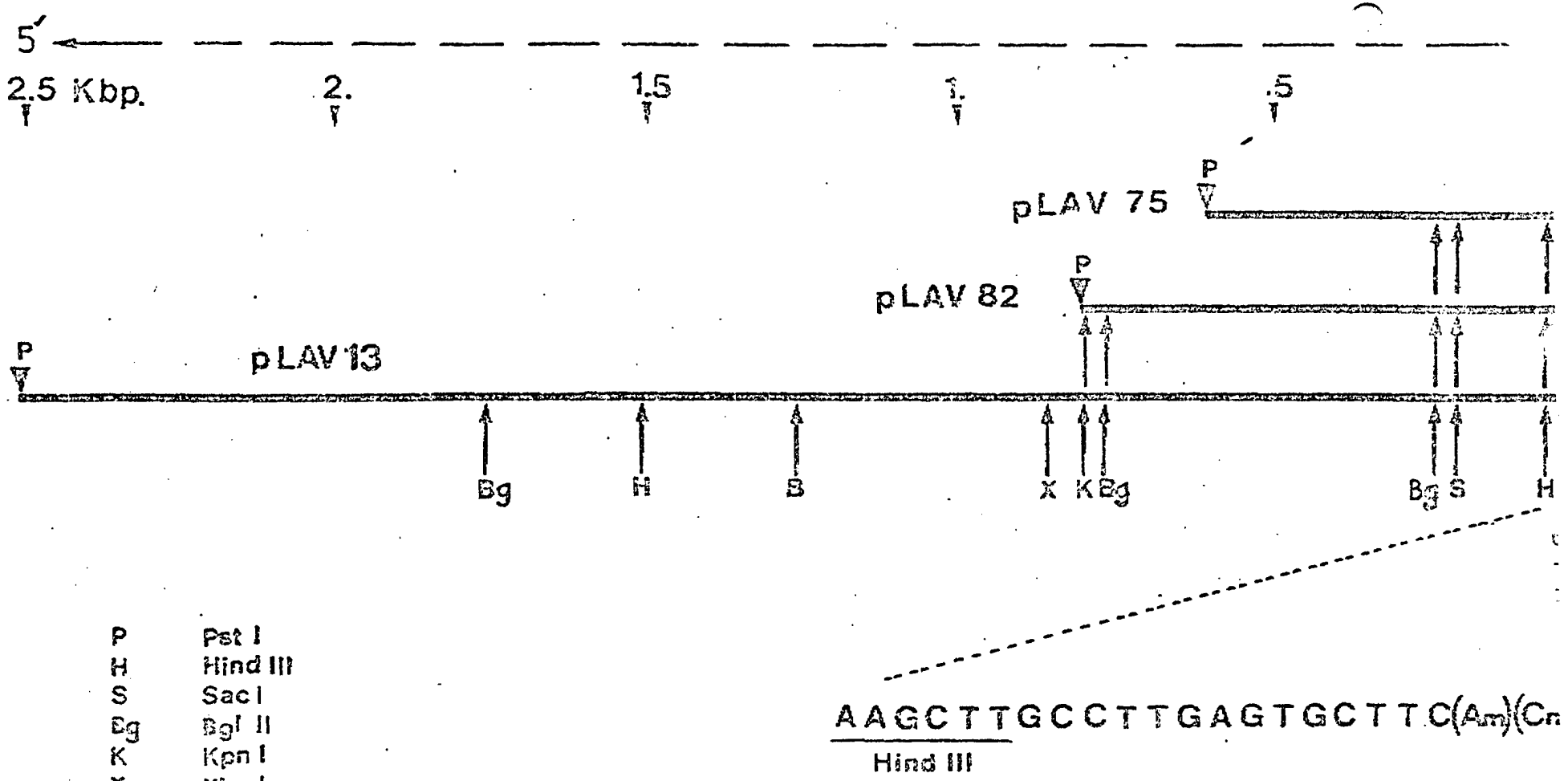
5'
2.5 Kbp.
2.
1.5
1.
.5
P
pLAV 75
P
pLAV 82
P
pLAV 13
Bg
H
B
X
K
Bg
Bg
S
H
P Pst I
H Hind III
S Sac I
Bg Bgl II
K Kpn I
X Xho I
B BamHI
AAGCTTGCCTTGAGTGCTTC(Am)(Cn
Hind III

FIG.1.

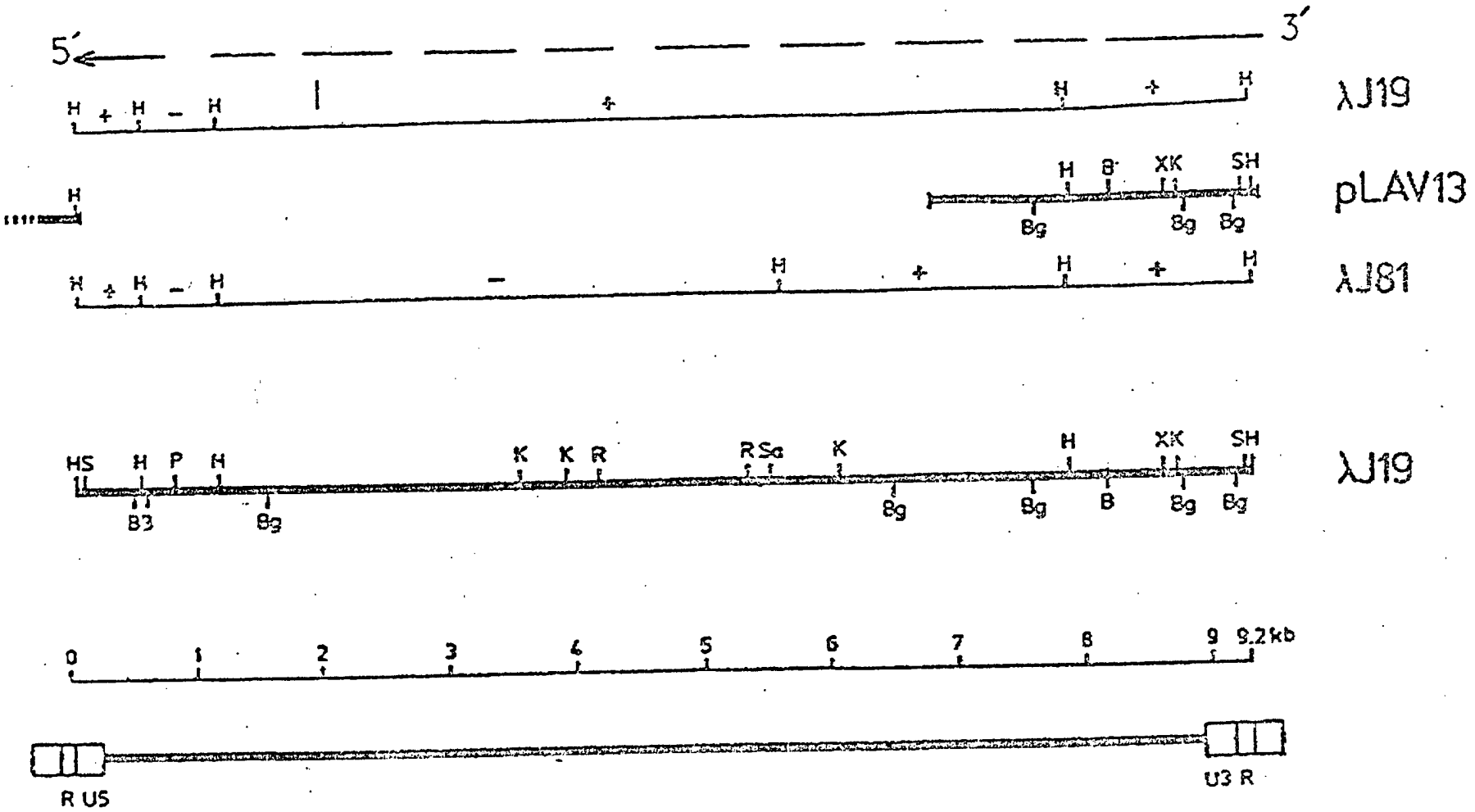

FIG.2.

H= Hind III K_Kpn I
S= Sac I R_EcoRI
B= Bam HI Sa_Sal I
P= Pst I. X _XhoI
Bg=Bgl II